

FILE DEFINITION TEST CHANGE

Notification Date: April 22, 2025 Effective Date: May 22, 2025

Hypobetalipoproteinemia Gene Panel, Varies

Test ID: HYPBG

Explanation: On the effective date, the test algorithm will be updated to include a reflex test when necessary. Additionally, the Ordering Guidance, Specimen Required and Report Available sections will be updated as indicated below.

New Reflex Tests				
Test ID	Reporting Name	Available Separately	Always Performed	
CULFB	Fibroblast Culture for Genetic Test	Yes	No	

Current Testing Algorithm				
None				

New Testing Algorithm

Skin biopsy: For skin biopsy or cultured fibroblast specimens, a fibroblast culture will be performed at an additional charge. If viable cells are not obtained, the client will be notified.

Current Ordering Guidance

This gene panel contains genes in common with HCHLG / Hypercholesterolemia Gene Panel, Varies. These tests should not be ordered concurrently as they assist in diagnosing conflicting disorders. For low levels of cholesterol (hypocholesterolemia), order this test. For high levels of cholesterol (hypercholesterolemia), order HCHLG.

Customization of this panel and single gene analysis for any gene present on this panel are available. For more information see CGPH / Custom Gene Panel, Hereditary, Next-Generation Sequencing, Varies.

New Ordering Guidance

This gene panel contains genes in common with HCHLG / Hypercholesterolemia Gene Panel, Varies. These tests should not be ordered concurrently as they assist in diagnosing conflicting disorders. For low levels of cholesterol (hypocholesterolemia), order this test. For high levels of cholesterol (hypercholesterolemia), order HCHLG.

Customization of this panel and single gene analysis for any gene present on this panel are available. For more information see CGPH / Custom Gene Panel, Hereditary, Next-Generation Sequencing, Varies. To modify this panel via CGPH, use the Cardiovascular/Connective Tissue/Dyslipidemia/Cerebrovascular/Primary Ciliary Dyskinesia disease state for step 1 on the Custom Gene Ordering Tool.

Targeted testing for familial variants (also called site-specific or known mutations testing) is available for the genes on this panel. See FMTT / Familial Variant, Targeted Testing, Varies. To obtain more information about this testing option, call 800-533-1710.

Targeted testing for familial variants (also called site-specific or known mutations testing) is available for the genes on this panel. See FMTT / Familial Variant, Targeted Testing, Varies. To obtain more information about this testing option, call 800-533-1710.

Current Specimen Required

Patient Preparation: A previous bone marrow transplant from an allogenic donor will interfere with testing. Call 800-533-1710 for instructions for testing patients who have received a bone marrow transplant.

Specimen Type: Whole blood

Container/Tube:

Preferred: Lavender top (EDTA) or yellow top

(ACD)

Acceptable: Any anticoagulant Specimen Volume: 3 mL Collection Instructions:

- 1. Invert several times to mix blood
- 2. Send whole blood specimen in original tube. Do not aliquot.

Specimen Stability Information: Ambient

(preferred)/Refrigerated

New Specimen Required

Patient Preparation: A previous hematopoietic stem cell transplant from an allogenic donor will interfere with testing. For instructions for testing patients who have received a hematopoietic stem cell transplant, call 800-533-1710.

Submit only 1 of the following specimens:

Specimen Type: Whole blood

Container/Tube:

Preferred: Lavender top (EDTA) or yellow top (ACD)

Acceptable: Green top (Sodium heparin)

Specimen Volume: 3 mL Collection Instructions:

- 1. Invert several times to mix blood.
- 2. Send whole blood specimen in original tube. Do not aliquot.

Specimen Stability Information: Ambient (preferred) 4 days/Refrigerated 4 days/Frozen 4 days

Additional Information:

- 1. Specimens are preferred to be received within 4 days of collection. Extraction will be attempted for specimens received after 4 days, and DNA yield will be evaluated to determine if testing may proceed.
- 2. To ensure minimum volume and concentration of DNA is met, the requested volume must be submitted. Testing may be canceled if DNA requirements are inadequate.

Specimen Type: Cord blood

Container/Tube:

Preferred: Lavender top (EDTA) or yellow top (ACD)

Acceptable: Green top (Sodium heparin)

Specimen Volume: 3 mL Collection Instructions:

- 1. Invert several times to mix blood.
- 2. Send cord blood specimen in original tube. Do not aliquot. **Specimen Stability Information:** Ambient (preferred) 4

days/Refrigerated 4 days/Frozen 4 days

Additional Information:

- 1. Specimens are preferred to be received within 4 days of collection. Extraction will be attempted for specimens received after 4 days, and DNA yield will be evaluated to determine if testing may proceed.
- 2. To ensure minimum volume and concentration of DNA is met, the requested volume must be submitted. Testing may be canceled if DNA requirements are inadequate.
- 3. While a properly collected cord blood sample may not be at risk for maternal cell contamination, unanticipated complications may occur during collection. Therefore,

maternal cell contamination studies are recommended to ensure the test results reflect that of the patient tested and are available at an additional charge. Order MATCC / Maternal Cell Contamination, Molecular Analysis, Varies on the maternal specimen.

Specimen Type: Saliva

Patient Preparation: Patient should not eat, drink, smoke, or

chew gum 30 minutes prior to collection. **Supplies:** Saliva Collection Kit (T786)

Specimen Volume: 2 Swabs
Collection Instructions: Collect and send specimen per kit

instructions.

Specimen Stability Information: Ambient (preferred) 30

days/Refrigerated 30 days

Additional information: Saliva specimens are acceptable but not recommended. Due to lower quantity/quality of DNA yielded from saliva, some aspects of the test may not perform as well as DNA extracted from a whole blood sample. When applicable, specific gene regions that were unable to be interrogated will be noted in the report. Alternatively, additional specimen may be required to complete testing.

Specimen Type: Cultured fibroblasts

Source: Skin

Container/Tube: T-25 flask Specimen Volume: 2 Flasks

Collection Instructions: Submit confluent cultured fibroblast cells from a skin biopsy. Cultured cells from a prenatal

specimen will not be accepted.

Specimen Stability Information: Ambient (preferred) <24

hours/Refrigerated <24 hours

Additional Information:

- 1. Specimens are preferred to be received within 24 hours of collection. Culture and extraction will be attempted for specimens received after 24 hours and will be evaluated to determine if testing may proceed.
- A separate culture charge will be assessed under CULFB / Fibroblast Culture for Biochemical or Molecular Testing. An additional 3 to 4 weeks are required to culture fibroblasts before genetic testing can occur.

Specimen Type: Tissue biopsy Supplies: Hank's Solution (T132)

Container/Tube: Sterile container with sterile Hank's balanced salt solution, Ringer's solution, or normal saline.

Specimen Volume: 0.5 to 3 cm(3) or larger

Specimen Stability Information: Ambient (preferred) <24

hours/Refrigerated <24 hours

Additional Information:

- 1. Specimens are preferred to be received within 24 hours of collection. Culture and extraction will be attempted for specimens received after 24 hours and will be evaluated to determine if testing may proceed.
- A separate culture charge will be assessed under CULFB / Fibroblast Culture for Biochemical or Molecular Testing. An additional 3 to 4 weeks are required to culture fibroblasts before genetic testing can occur.

Specimen Type: Blood spot

Supplies: Card-Blood Spot Collection (Filter Paper) (T493)

Container/Tube:

Preferred: Collection card (Whatman Protein Saver 903

Paper)

Acceptable: PerkinElmer 226 filter paper or blood spot

collection card

Specimen Volume: 2 to 5 Blood spots

Collection Instructions:

- 1. An alternative blood collection option for a patient older than 1 year is a fingerstick. For detailed instructions, see How to Collect a Dried Blood Spot Sample.
- 2. Let blood dry on the filter paper at ambient temperature in a horizontal position for a minimum of 3 hours.
- 3. Do not expose specimen to heat or direct sunlight.
- 4. Do not stack wet specimens.
- 5. Keep specimen dry.

Specimen Stability Information: Ambient (preferred)/Refrigerated

Additional Information:

- 1. Blood spot specimens are acceptable but not recommended. Due to lower quantity/quality of DNA yielded from blood spots, some aspects of the test may not perform as well as DNA extracted from a whole blood sample. When applicable, specific gene regions that were unable to be interrogated will be noted in the report. Alternatively, additional specimen may be required to complete testing.
- 2. Due to lower concentration of DNA yielded from blood spot, it is possible that additional specimen may be required to complete testing.
- 3. For collection instructions, see <u>Blood Spot Collection</u> Instructions
- 4. For collection instructions in Spanish, see <u>Blood Spot</u> Collection Card-Spanish Instructions (T777)
- 5. For collection instructions in Chinese, see <u>Blood Spot Collection Card-Chinese Instructions</u> (T800)

Specimen Type: Extracted DNA

Container/Tube:

Preferred: Screw Cap Micro Tube, 2 mL with skirted conical

base

Acceptable: Matrix tube, 1 mL Collection Instructions:

- 1. The preferred volume is at least 100 mcL at a concentration of 75 ng/mcL.
- 2. Include concentration and volume on tube.

Specimen Stability Information: Frozen (preferred) 1

year/Ambient/Refrigerated

Additional Information: DNA must be extracted in a CLIA-certified laboratory or equivalent and must be extracted from a specimen type listed as acceptable for this test (including applicable anticoagulants). Our laboratory has experience with Chemagic, Puregene, Autopure, MagnaPure, and EZ1 extraction platforms and cannot guarantee that all extraction methods are compatible with this test. If testing fails, one repeat will be attempted, and if unsuccessful, the test will be reported as failed and a charge will be applied. If applicable, specific gene regions that were unable to be interrogated due to DNA quality will be noted in the report.

Current Report Available	New Report Available
28 to 42 days	21 to 28 days

Questions

Contact Michelle Raths, Laboratory Resource Coordinator at 800-533-1710.